

Structure of Hormaomycin, a Naturally Occurring Cyclic Octadepsipeptide, in the Crystal

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The structure of hormaomycin has been determined in two crystals grown under different conditions, *i. e.* in the absence and in the presence of magnesium chloride. In both crystals, the macrocyclic hexadepsipeptide assumes a rather flat conformation, and the dipeptide side chain resides in the same equatorial plane. This is a significant difference in comparison with the compact bent conformation of hormaomycin in solution, as previously determined by an extensive NMR study.

Key words: Crystal Structure, Hormaomycin, Macrocyclic Depsipeptide, X-Ray Diffraction

Introduction

Hormaomycin **1**, a macrocyclic depsipeptide consisting of eight amino acid residues (Fig. 1), was first isolated and structurally elucidated in 1989–1990 [1, 2]. Its absolute configuration was fully assigned some 15 years later [3, 4] and rigorously confirmed by total synthesis [5].

Since hormaomycin **1** features some interesting biological activity [1, 2] including a significant inhibitory effect against *Plasmodium falciparum*, the pathogen causing malaria [6], more than 20 analogs were prepared to enable, among others, the study of structure-activity relationships and the conformational behavior in solution [7–9]. Detailed NMR-spectroscopic analysis revealed a rather compact secondary structure of the octapeptide, and it appeared that this compact conformation is essential for the biological activity [9]. It would certainly be interesting to know whether that same conformation prevails in the crystal. Several earlier attempts to crystallize the native hormaomycin were not successful, but finally crystals of the synthetic material **1** could be obtained and subjected to X-ray diffraction.

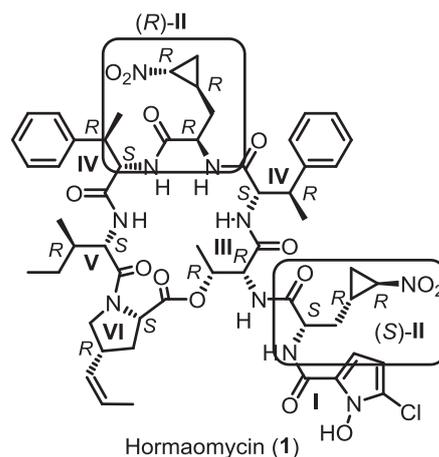


Fig. 1. Structural formula with absolute configuration of hormaomycin (**1**). I: Chpca; II: (1R',2R')-(3-Ncp)Ala; III: (2R)-α-Thr; IV: (2S,3R)-(βMe)Phe; V: (S)-Ile; VI: (2S,4R)-4-(Z)-(4-PE)Pro.

Results and Discussion

Hormaomycin crystallized in space groups *P1* and *P2*₁ from aqueous solutions at weakly basic pH = 8.5,

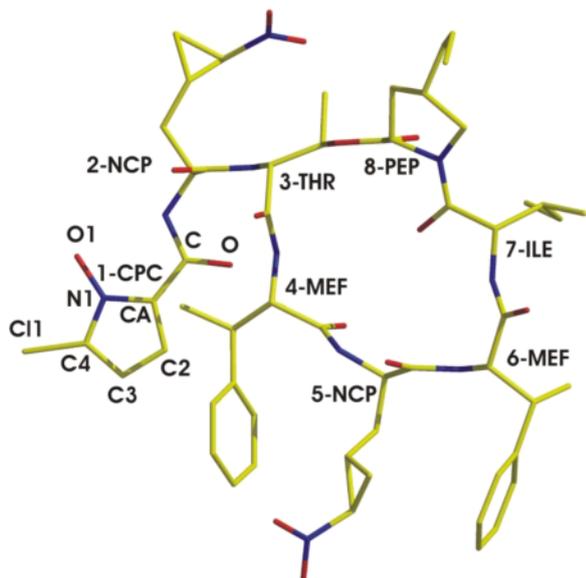


Fig. 2 (color online). Monomer structure of hormaomycin with 3-letter residue labels which correspond to the naming scheme in the deposited CIF files. CPC: **I** (Chpca); 2-NCP: (*S*)-**II** [(2*S*,1*R*',2*R*')-(3-Ncp)Ala]; 5-NCP: (*R*)-**II** [(2*R*,1*R*',2*R*')-(3-Ncp)Ala]; THR: **III** [(2*R*)- α -Thr]; MEF: **IV** [(2*S*,3*R*)-(β Me)Phe]; ILE: **VI**: [(2*S*,4*R*)-4-(*Z*)-(4-PE)Pro]. Only the atom names for Chpca are given explicitly. For better clarity other atom names are not explicitly given. However, the atom names follow the usual amino acid naming schemes.

depending on the presence or absence of MgCl_2 , respectively. Both crystals contain two independent molecules. The monomer structure of hormaomycin (**1**) is shown in Fig. 2, while Fig. 3 displays the asymmetric unit of the structure in space group *P1* with a six-fold water-coordinated Mg^{2+} ion. All molecules assume an almost planar conformation and share high local homology. The root-mean-square deviation (rmsd) for the superposition between one molecule from each crystal form is 1.4 Å for 118 atoms. The strongest deviations occur in the flexible side chains, and the rmsd only for the main chain atoms of the flat ring section is 0.26 Å for 24 atoms. In contrast to the crystal structure, the structure in CDCl_3 solution, as previously determined in an elaborate NMR-spectroscopic study [9], shows a compactly folded conformation, in which the residues **I** (Chpca) and (*S*)-**II** [(2*S*,1*R*',2*R*')-(3-Ncp)Ala] are bent almost perpendicular to the rather flat main ring consisting of residues **III**–**VI**, as illustrated in Fig. 4. This

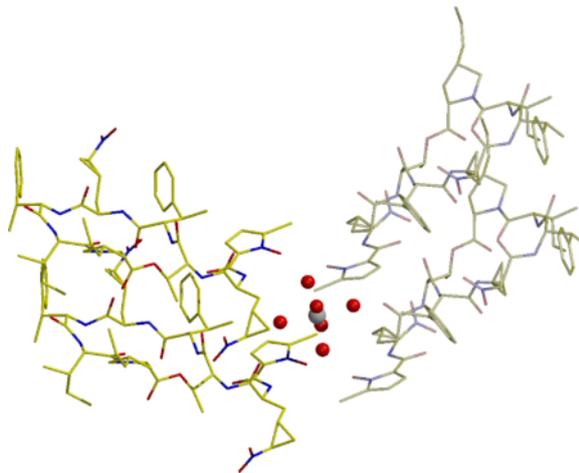


Fig. 3 (color online). Hormaomycin (**1**) in space group *P1* with a Mg^{2+} ion in six-fold coordination with water molecules surrounded by four hormaomycin molecules.

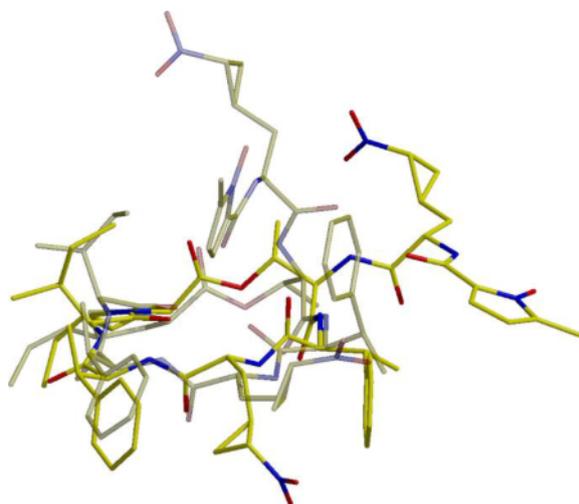


Fig. 4 (color online). Superposition of the *P1* crystal structure (yellow carbon atoms) with the NMR-derived structure in solution (semi-transparent). The latter assumes a more compact conformation with residues (*S*)-**II** [(2*S*,1*R*',2*R*')-(3-Ncp)Ala] and **III** [(2*R*)- α -Thr], and the side chains of **IV** [(2*S*,3*R*)-(β Me)Phe] and **V** [(*S*)-Ile] being bent towards the center of the rather flat main ring.

most probably is caused by an energetically favorable π -stacking interaction between the *N*-hydroxy-2-chloropyrrol moiety at the end of the side-chain **I** (Chpca) and one of the phenyl groups of the two β -methylphenylalanine residues **IV** in the macrocycle. In DMSO solution, however, the π -stacking interaction

Table 1. Backbone torsion angles for all four independent crystal structures of hormaomycin in comparison with those obtained by NMR spectroscopy. Torsion angle definitions: ϕ : $C_{i-1}-N_i-C_{\alpha,i}-C_i$; ψ : $N_{i-1}-C_{\alpha,i-1}-C_{i-1}-N_i$; ω : $C_{\alpha,i-1}-C_{i-1}-N_i-C_{\alpha,i}$.

| Building block | Torsion angle ϕ (deg) | | | NMR ^a | |
|--|----------------------------|--------------------------------|------------|------------------|-------------------|
| | In <i>P1</i> form | In <i>P2</i> ₁ form | Mean value | DMSO | CDCl ₃ |
| I (Chpca) | – ^b | – | – | – | – |
| (<i>S</i>)- II [(2 <i>S</i> ,1 <i>R'</i> ,2 <i>R'</i>)-(3-Ncp)Ala] | –71, –70 | –57, –65 | –65.8 | * ^c | –68 |
| (<i>R</i>)- II [(2 <i>R</i> ,1 <i>R'</i> ,2 <i>R'</i>)-(3-Ncp)Ala] | 140, 137 | 124, 131 | 133.0 | 116 | 69 |
| III [(2 <i>R</i>)- α -Thr] | 76, 78 | 63, 60 | 69.3 | * ^c | 100 |
| IVa [(2 <i>S</i> ,3 <i>R</i>)-(β Me)Phe] | –122, –120 | –99, –104 | –111.3 | –98 | –67 |
| IVb [(2 <i>S</i> ,3 <i>R</i>)-(β Me)Phe] | –92, –89 | –79, –79 | –84.8 | –83 | –90 |
| V (<i>S</i>)-Ile | –138, –132 | –109, –110 | –122.3 | –128 | –93 |
| VI (2 <i>S</i> ,4 <i>R</i>)-4-(<i>Z</i>)-(4-PE)Pro | –62, –60 | –58, –61 | –60.3 | –66 | –61 |

| Building block | Torsion angle ψ (deg) | | | NMR ^a | |
|--|----------------------------|--------------------------------|------------|------------------|-------------------|
| | In <i>P1</i> form | In <i>P2</i> ₁ form | Mean value | DMSO | CDCl ₃ |
| I (Chpca) | – ^b | – | – | – | – |
| (<i>S</i>)- II [(2 <i>S</i> ,1 <i>R'</i> ,2 <i>R'</i>)-(3-Ncp)Ala] | 123, 122 | 107, 115 | 116.8 | * ^c | 105 |
| (<i>R</i>)- II [(2 <i>R</i> ,1 <i>R'</i> ,2 <i>R'</i>)-(3-Ncp)Ala] | –150, –154 | –161, –164 | –157.3 | –133 | –135 |
| III [(2 <i>R</i>)- α -Thr] | 36, 33 | 28, 29 | 31.5 | * ^c | –69 |
| IVa [(2 <i>S</i> ,3 <i>R</i>)-(β Me)Phe] | 132, 133 | 135, 133 | 133.3 | 126 | 180 |
| IVb [(2 <i>S</i> ,3 <i>R</i>)-(β Me)Phe] | –3, –3 | –20, –16 | –10.5 | –7 | –47 |
| V (<i>S</i>)-Ile | 173, 169 | 165, 167 | 168.5 | 160 | 152 |
| VI (2 <i>S</i> ,4 <i>R</i>)-4-(<i>Z</i>)-(4-PE)Pro | – ^b | – | – | – | – |

| Building block | Torsion angle ω (deg) | | | NMR ^a | |
|--|------------------------------|--------------------------------|------------|------------------|-------------------|
| | In <i>P1</i> form | In <i>P2</i> ₁ form | Mean value | DMSO | CDCl ₃ |
| I (Chpca) | 176, 173 | 183, 187 | 179.8 | * ^c | 182 |
| (<i>S</i>)- II [(2 <i>S</i> ,1 <i>R'</i> ,2 <i>R'</i>)-(3-Ncp)Ala] | 178, 181 | 181, 179 | 179.8 | * ^c | 178 |
| (<i>R</i>)- II [(2 <i>R</i> ,1 <i>R'</i> ,2 <i>R'</i>)-(3-Ncp)Ala] | 187, 187 | 184, 181 | 184.8 | 178 | 192 |
| III [(2 <i>R</i>)- α -Thr] | 185, 184 | 180, 180 | 182.3 | –173 | 170 |
| IVa [(2 <i>S</i> ,3 <i>R</i>)-(β Me)Phe] | 180, 178 | 171, 173 | 175.5 | 170 | 166 |
| IVb [(2 <i>S</i> ,3 <i>R</i>)-(β Me)Phe] | 169, 168 | 173, 171 | 170.3 | 180 | 173 |
| V (<i>S</i>)-Ile | 181, 188 | 183, 182 | 183.5 | * ^c | 184 |
| VI (2 <i>S</i> ,4 <i>R</i>)-4-(<i>Z</i>)-(4-PE)Pro | – ^b | – | – | – | – |

^a As reported for the NMR models in DMSO [10] and CDCl₃ [9]; ^b empty entries mark undefined angles in the respective residue; ^c not reported in ref. [10].

apparently does not play a significant role, and the conformation of the side chain is less well defined [10]. Table 1 compares the mean backbone angles of the two crystallographic structures with those reported by Blackledge and Griesinger [10] for the NMR models in DMSO and CDCl₃. As was assumed for the NMR-derived structures, in the crystals all peptide bonds have an *s-trans* orientation.

Conclusion

Two different crystal structures of hormaomycin (**1**) with two independent molecules each assume an open, flat conformation and show no *s-cis*-peptide bonds. The overall similarity of all four molecules is very

high with small variations in the flexible side chains. This structure in the crystal significantly differs from the compact folded structure in CDCl₃ solution [9], but in DMSO the side chain is more or less in a similar orientation as in the crystal. Blackledge, Griesinger *et al.* reported rms deviations between the superimposed macrocyclic rings of 2.3 Å between the solution and crystal structures, but only of 0.66 Å between the structures in the two different solvents [10]. They attributed these differences to the different hydrogen-bonding properties of the solvents involved. In view of the much larger differences between the crystal and the NMR-derived structures than between the latter two, we suggest that intermolecular interactions in the crystal may have even more influence on the confor-

mations than the solvents. Since hormaomycin in this study crystallized under aqueous conditions, we suggest that the crystal structures represent the conformation of hormaomycin *in vivo*. Whereas at least one of the NMR studies [10] appears to have used natural hormaomycin, the crystal structure analyses were performed on the synthetic product. Since these atomic resolution structures were solved by *ab initio* Direct Methods without making any assumptions about the chemical structures involved, they provide a convincing confirmation that the natural and synthetic compounds are indeed identical.

Experimental Section

Crystals were grown by the hanging drop vapor diffusion method. A drop (1 μL) of a solution of approx. 56 g L^{-1} (*w/v*) hormaomycin (**1**) in EtOH mixed with 1 μL of an aqueous buffer solution was hung over 300 μL of the reservoir solution. The reservoir solution for the crystal in space group $P2_1$ contained 0.1 M Tris-HCl (2-amino-2-

hydroxymethylpropane-1,3-diol adjusted to pH = 8.5 by addition of aq. HCl solution), 50% MPD (2-methylpentane-2,4-diol) and 5% dioxane, while the reservoir solution for the crystal in space group $P1$ contained 0.1 M Tris-HCl (pH = 8.5), 50% isopropanol and 20 mM MgCl_2 . The crystal form $P2_1$ was improved by micro-seeding. X-ray data were collected at the synchrotrons DESY in Hamburg, Germany ($P2_1$), and at the Paul Scherrer Institute (Swiss Light Source) in Switzerland ($P1$). The crystals diffracted poorly even with synchrotron radiation, so although the structures were unambiguously determined, their precision is not as high as usual for small molecule crystal structures. Data were processed and scaled with XDS [11]. The structures were solved and refined with SHELX-2013 [12]. The models were built with COOT [13] and SHELXLE [14]. Restraints for solvent molecules were generated from the grade-server (<http://grade.globalphasing.org>). Figures were created with MOLSCRIPT [15] and RASTER3D [16].

Refinement of $\mathbf{1} \times [\text{Mg}(\text{H}_2\text{O})_6]\text{X}_2$ (space group $P1$): Due to the poor quality of the intensity data set the Flack parameter could not be determined reliably. The chirality was determined relative to the chirality of the standard L-amino acids.

Table 2. Crystal structure data for $\mathbf{1} \times [\text{Mg}(\text{H}_2\text{O})_6]\text{X}_2$ and **1**.

| | $\mathbf{1} \times [\text{Mg}(\text{H}_2\text{O})_6]\text{X}_2$ | 1 |
|---|---|---|
| Formula | ($\text{C}_{55}\text{H}_{68}\text{ClN}_{10}\text{O}_{14}$) ₂ , ($\text{C}_3\text{H}_7\text{O}$) ₂ , Mg, O ₁₈ $\text{C}_{116}\text{H}_{150}\text{Cl}_2\text{MgN}_{20}\text{O}_{48}$ | ($\text{C}_{55}\text{H}_{68}\text{N}_{10}\text{O}_{14}\text{Cl}$) ₂ , ($\text{C}_6\text{H}_{12}\text{O}_2$) ₄ , ($\text{C}_4\text{H}_8\text{O}_2$) ₂ , Na, O ₁₀ $\text{C}_{142}\text{H}_{200}\text{Cl}_2\text{N}_{20}\text{NaO}_{50}$ |
| M_r | 2687.76 | 3081.10 |
| Cryst. size (max/mid/min), mm | 0.10/0.20/0.20 | 0.07/0.07/0.05 |
| Crystal system | triclinic | monoclinic |
| Space group | $P1$ (no. 1) | $P2_1$ (no. 4) |
| Wavelength, Å | 0.77492 | 0.8430 |
| a , Å | 7.370(2) | 14.080(3) |
| b , Å | 15.969(4) | 32.760(7) |
| c , Å | 30.280(5) | 18.660(4) |
| α , deg | 92.086(10) | 90 |
| β , deg | 91.01(2) | 101.73(3) |
| γ , deg | 102.693(10) | 90 |
| V , Å ³ | 3473.1(13) | 8427(3) |
| Z | 1 | 2 |
| T , K | 100(2) | 100(2) |
| $D_{\text{calcd.}}$, g cm^{-3} | 1.29 | 1.21 |
| μ (MoK α), cm^{-1} | 0.2 | 0.2 |
| $F(000)$, e | 1416 | 3274 |
| hkl range | $\pm 7, \pm 17, \pm 31$ | $-12/13, \pm 31, -16/18$ |
| θ (min/max), deg | 1.63/27.99 | 1.48/24.40 |
| Refl. measured | 24 167 | 28 028 |
| Refl. unique/ R_{int} | 16 630/0.0943 | 14 773/0.0844 |
| Refl with $I > 2 \sigma(I)$ | 14 734 | 10 235 |
| Data/restraints/parameters | 16 630/2717/1589 | 14 773/3784/1921 |
| $R(F)/wR(F^2)$ ^a (all refls.) | 0.1313/0.3267 | 0.1632/0.3527 |
| GoF (F^2) ^b | 1.312 | 1.216 |
| $\Delta\rho_{\text{fin}}$ (max/min), $e \text{ \AA}^{-3}$ | 1.05/−0.75 | 0.68/−0.47 |

Only the two antibiotics molecules and the Mg with its six coordinating oxygen atoms were refined anisotropically, and no attempt was made to place hydrogens on the water oxygens.

Refinement of **1** (space group $P2_1$): As above, the chirality was determined relative to the chirality of the standard L-amino acids. Only the two antibiotics molecules were refined anisotropically, and no attempt was made to place hydrogens on the water oxygens. Despite the low density and high U values for MPD (2-methyl-2,4-pentanediol) and DOX (dioxane), they show up as difference density when the molecules are removed. The large shift/error and the short H...H contacts reported by PLATON refer to one of the dioxane molecules. The data resolution does not allow the detailed identification of ions nor a discussion of hydrogen bonds.

Table 2 contains the crystal data and numbers pertinent to data collection and structure refinement of both structure determinations.

CCDC 1003606 contains the supplementary crystallographic data for space group $P1$. CCDC 1003607 contains the supplementary crystallographic data for space group $P2_1$. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

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- [1] N. Andres, H. Wolf, H. Zähler, E. Rössner, A. Zeeck, W. A. König, V. Sinnwell, *Helv. Chim. Acta* **1989**, *72*, 426–437.
- [2] E. Rössner, A. Zeeck, W. A. König, *Angew. Chem., Int. Ed. Engl.* **1990**, *29*, 64–65.
- [3] J. Zindel, A. Zeeck, W. A. König, A. de Meijere, *Tetrahedron Lett.* **1993**, *34*, 1917–1920.
- [4] B. D. Zlatopolskiy, K. Loscha, P. Alvermann, S. I. Kozhushkov, S. V. Nikolaev, A. Zeeck, A. de Meijere, *Chem. Eur. J.* **2004**, *10*, 4708–4717.
- [5] B. D. Zlatopolskiy, A. de Meijere, *Chem. Eur. J.* **2004**, *10*, 4718–4727.
- [6] K. Otaguro, H. Ui, A. Ishiyama, N. Arai, M. Kobayashi, Y. Takahashi, R. Masumo, K. Shiomi, H. Yamada, S. Omura, *J. Antibiotics* **2003**, *56*, 322–324.
- [7] S. I. Kozhushkov, B. D. Zlatopolski, M. Brandl, P. Alvermann, M. Radzom, B. Geers, A. de Meijere, A. Zeeck, *Eur. J. Org. Chem.* **2005**, 854–863.
- [8] B. D. Zlatopolski, M. Radzom, A. Zeeck, A. de Meijere, *Eur. J. Org. Chem.* **2006**, 1525–1534.
- [9] U. M. Reinscheid, B. D. Zlatopolskiy, C. Griesinger, A. Zeeck, A. de Meijere, *Chem. Eur. J.* **2005**, *11*, 2929–2945.
- [10] U. M. Reinscheid, J. Farjon, M. Radzom, P. Haberk, A. Zeeck, M. Blackledge, C. Griesinger, *ChemBioChem* **2006**, *7*, 287–296.
- [11] W. Kabsch, *Acta Crystallogr.* **2010**, *D66*, 125–132.
- [12] G. M. Sheldrick, *Acta Crystallogr.* **2008**, *A64*, 112–122.
- [13] P. Emsley, B. Lohkamp, W. Scott, K. Cowtan, *Acta Crystallogr.* **2010**, *D66*, 486–501.
- [14] C. B. Hübschle, G. M. Sheldrick, B. Dittrich, *J. Appl. Crystallogr.* **2011**, *44*, 1281–1284.
- [15] P. J. Kraulis, *J. Appl. Crystallogr.* **1991**, *24*, 946–950.
- [16] E. A. Merritt, D. J. Bacon, *Methods Enzymol.* **1997**, *277*, 505–524.