Introduction

Astaxanthin (3,3'-dihydroxy-β,β'-carotene-4,4'-dione), a particular carotenoid, is widely used as food colourant in aquaculture industries. Since the animals involved are unable to synthesize astaxanthin, this compound has to be supplemented to the feed to ensure proper pigmentation like their wild brethren (Tejera et al., 2007).

Moreover, there has recently been interest in the medical properties of astaxanthin. Many studies have demonstrated that astaxanthin may possess powerful antioxidant activity, which may be ten times stronger than that of other carotenoids such as β-carotene, zeaxanthin, and canthaxanthin (Nakano et al., 1999). Therefore, astaxanthin exerts a protective effect against chronic diseases such as cancer and other degenerative diseases (Hussein et al., 2006).

The red yeast Xanthophyllomyces dendrorhous (previously named Phaffia rhodozyma, a moderately psychrophilic yeast) has been regarded as one of the promising microbiological production systems for natural astaxanthin (Dominguez-Bocanegra et al., 2007). In X. dendrorhous, astaxanthin represents about 85% of the total carotenoids content. Considerable research efforts have been devoted to improve the astaxanthin production. The production of astaxanthin in X. dendrorhous can be increased by classical mutagenesis (An et al., 1989; Bon et al., 1997). In our laboratory, X. dendrorhous strains were successively treated with UV radiation and mutagenic chemicals, and then screened for high-yield mutants. During that process, we isolated more than twenty mutants that produced more astaxanthin, but their colony's colour was obviously different, indicating different accumulation of carotenoids intermediates. In order to understand the changes in the carotenoids' synthesis pathway, it would be valuable to develop a reliable and applicable method to characterize the carotenoid profile of X. dendrorhous. Preferably, the method should be able to identify and quantify astaxanthin and other carotenoids simultaneously.

Over the years, high-performance liquid chromatography (HPLC) has been widely applied in carotenoids analysis using isocratic or gradient mobile phases in either normal-phase or reversed-phase mode (Su et al., 2002). Rao et al. (2005) isolated and characterized some minor impurities of astaxanthin using a normal-phase silica gel column with n-hexane/acetone/tetrahydrofuran as mobile phase and identified related impurities by EMS and NMR spectroscopy. Lin et al. (2005) developed a reversed-phase HPLC method to analyse carotenoids in spear shrimp shells. Reversed-phase liquid chromatography with typical eluents, including methanol, acetonitrile, and water, was
used for separation of carotenoids in a variety of biological materials (Su et al., 2002; Verdoes et al., 2003). Weber et al. (2007) developed a reversed-phase HPLC method with acetone/water as solvent system to analyse and identify some carotenoids by liquid chromatography-atmospheric pressure chemical ionization-mass spectrometry (LC-APCI-MS). However, it was often found that astaxanthin failed to yield a symmetrical peak, and the carotenoids peaks’ resolution was poor. Abu-Lafi and Turujman (1999) have used a Pirkle (Regis, USA) covalent L-leucine column for rapid and direct resolution of the stereoisomers of all-trans-astaxanthin, but the reproducibility of this method was found to be poor when repeated with an identical L-leucine column. Subsequently, a reversed-phase HPLC method based on a C30 column was developed for separation of cis/trans isomers of different carotenoids (Su et al., 2002; Dugo et al., 2008). However, a C30 column is not suitable for routine analyses because of its high cost. So far, there are no reports on a method for simultaneous isolation and characterization of astaxanthin and carotenoids from X. dendrorhous.

The purpose of the present study was to develop and validate a method for simultaneous analysis of astaxanthin and its carotenoid precursors from X. dendrorhous fermentation using C18 reversed-phase HPLC.

Material and Methods

Strains and chemicals

X. dendrorhous strain AS 2.1557 was obtained from China General Microbiological Culture Collection Center (CGMCCC, Beijing, China). It was subjected to successive mutagenesis according to the method described by An et al. (1989). Mutants were maintained at −70 °C in yeast malt (YM) broth and 30% glycerol. In this work, an astaxanthin-overproducing strain was used.

The shake flask medium was prepared in our laboratory; it was composed of 50 g glucose, 4 g yeast extract, 3 g KH2PO4, 3 g MgSO4 · 7H2O in 1 L distilled water, pH was adjusted to 5.5 using 1.0 m HCl. For solid medium, 2% agar was added. The yeast was incubated in 500-mL flasks containing 60 mL medium. Incubation was on an orbital shaker (200 rpm) for 5 d at 20 °C.

All reagents were of analytical reagent grade unless stated otherwise. HPLC grade solvents, including methanol, acetonitrile, and dichloromethane, were purchased from Merck, Germany. Silica gel G60 for thin-layer chromatography was obtained from Qingdao Haiyang chemical company, Qingdao City, China. Astaxanthin and β-carotene standard were purchased from Sigma-Aldrich, Shanghai, China. Ultra-pure water was prepared using a Milli-Q Academic A10 system (Millipore).

Sample preparation

Preparation of crude carotenoids from X. dendrorhous was carried out according to the literature (Weber et al., 2007) with minor modifications. In brief, 40 mL of culture broth were used. Cells were harvested by centrifugation. The obtained pellet was frozen at −20 °C, suspended in 3–5 mL DMSO by vortexing, and incubated at room temperature for 5 h. Following centrifugation and collection of the DMSO phase, the obtained pellet was suspended in 10 mL acetone with a spatula and by vortexing, and centrifuged. The acetone phase was decanted, and the DMSO and acetone extraction procedure was repeated further twice. All coloured DMSO and acetone phases were pooled and transferred to an extraction funnel containing an equal volume of light petroleum (b. p. range 30–75 °C), 10 mL distilled water and 5 mL saturated NaCl. Carotenoids were extracted into the light petroleum phase by gentle rotation, avoiding excessive agitation. The coloured light petroleum phase was washed three times with an equal volume of distilled water. NaCl was added, if required, for phase separation. The light petroleum phase was dried over Na2SO4 and rotary-evaporated to dryness at 40 °C in dim light.

The crude extract was taken up in acetone to 4.0 mL, giving a 10-fold volumetric concentration relative to the culture broth. Freezing was done at −20 °C to precipitate the colourless lipids. The defatted crude extracts were filtered through a 0.45-μm syringe filter, and 10-μL aliquots were subjected to reversed-phase HPLC analysis.

HPLC analysis

Analytical HPLC was performed with a Dikma (Tian Jin, China) Diamonsil TM-C18 column (5 μm; 250 mm × 4.6 mm) on a Waters 2695 series instrument equipped with an online diode-array detector. The mobile phase was filtered through a 0.45-μm membrane degassed before use. Under
isocratic conditions, the analysis was carried out at a flow rate of 1.0 mL/min at room temperature. Chromatograms were recorded at 480 nm, and UV-vis absorption spectra were recorded online with the photodiode-array detection system.

**LC-APCI-MS analysis**

In order to identify the main carotenoids appeared in the chromatogram, the mass spectra of carotenoids from *X. dendrorhous* yeast extracts...
were recorded. After DMSO/acetone extraction, the concentrated oil product was firstly chromatographed on activated silica gel TLC plates (silica gel G60, 5 x 20 cm). The solvent system used for TLC was acetone/n-hexane (1:5, v/v). The bands were scraped off and eluted with acetone. This TLC purification procedure was not required for HPLC analysis, but greatly improved the signal-to-noise ratio of mass spectra. LC-APCI-MS analysis was preformed as described (Weber et al., 2007). An Agilent LC-MSD-Trap-XCT instrument was operated at a fragmentor voltage of 140 V in the APCI-positive (PI) mode. The gas temperature was 350 °C, the vaporizer wasset to 400 °C; nitrogen was supplied as drying gas at 7 L/min, with a nebulizing pressure of 276 kPa. Spectra in the mass range between 100 and 800 were recorded. HPLC separation was carried out as described above.

**Results**

*Developing an HPLC method for determination of astaxanthin and carotenoids*

In order to develop a fine analysis method for astaxanthin and carotenoids, we choose reversed-phase HPLC using a C\textsubscript{18} column. Various mobile phases in isocratic or gradient mode were compared. Preliminary mobile phase studies were carried out with three solvents, viz., methanol, acetonitrile, and water in different compositions. The results showed that the peak resolution between astaxanthin and the other carotenoids was not satisfactory, and the astaxanthin peak was tailing. Then dichloromethane was added to the mobile phase; the quaternary mobile phase system provided better resolution than the previously used ternary solvent mixture. The most appropriate mobile phase was found to be composed of methanol/water/dichloromethane/acetonitrile (70:4:13:13, v/v/v/v). The resolution of peaks was improved. Most peaks were adequately resolved within 70 min. On the basis of the retention behaviour and absorption spectrum compared with the standards, peak 1 and peak 8 were identified as trans-astaxanthin and β-carotene, respectively. Astaxanthin showed a symmetric peak, and the resolution of the other carotenoids was excellent (Fig. 1a). The tolerance of the method was tested by adjusting the temperature and flow rate slightly. The separation between carotenoids was still fine (Figs. 1b, c).

**Method validation**

The results obtained showed good performance for astaxanthin analysis. So further studies on linearity and precision were performed.

The linearity of the standard curve was expressed in terms of the determination coefficient ($R^2$) from plots of the integrated peak area vs. concentration of the standard (in mg/L). A concentration range in concordance with the level of astaxanthin found in X. dendrorhous yeast extracts was tested. Results showed that the linearity from 50 mg/L to 250 mg/L was good (Fig. 2). Calibration lines were constructed, and showed linear correlation of $R^2 = 0.9978$, with a regression equation as follows: $y = 59463.50 + 58936.51x$.

The method precision, as repeatability, was evaluated on the basis of the relative standard deviation (RSD) of astaxanthin, with determination in 5 replicates of the same sample prepared from X. dendrorhous yeast. The results are shown in Table I. The RSD is 0.19% which is very good for routine analyses of astaxanthin.

**Related carotenoids identification by LC-APCI-MS**

Peak 1 and peak 8 were identified to be trans-astaxanthin and β-carotene, respectively, by spiking the sample with individual standards and comparing the retention times. The result was also verified by the LC-APCI mass spectrum. The other peaks were identified by LC-APCI-MS, UV-vis absorption and chemical characteristics. The molecular ions and the mass fragments
are listed in Table II. In the positive ion mode, the most prominent ion corresponded to [M+H]+ in the mass spectrum of carotenoids (Weber et al., 2007). The peaks 1, 2, 3, 4 and 5 had the fragments \([M+H−18]+\) and \([M+H−36]+\), which indicated that these carotenoids had two hydroxy groups (Ren and Zhang, 2008). The relative molecular weights of peak 3 and peak 4 were 596 according to \(m/z\) 597 [M+H]+, which were the same as that of trans-astaxanthin. These two peaks were identified by their online PDA UV-vis absorption maxima. Peak 3 was identified as 3,3’-dihydroxy-\(\beta\),\(\psi\)-carotene-4,4’-dione (DCD), which was first discovered by An et al. (1999) in the monocyclic carotenoid biosynthetic pathway. Peak 4 was identified as one or a mixture of astaxanthin isomers. Peak 5 showed the molecular weight of 594 and UV-vis spectra with absorption maxima at 461 and 429 nm. This absorption character was not found in the literature, but considering the existence of DCD in the system, we tentatively identified it as dehydro-DCD. Peak 6 and peak 7 were identified by their molecular weights and UV-vis spectra as torulene and \(\gamma\)-carotene, respectively.

**Discussion**

An earlier work using C\(_{18}\) reversed-phase HPLC to analyse carotenoids from yeasts has been reported in the literature. Weber et al. (2007) developed a C\(_{18}\) reversed-phase HPLC method by means of simple water/acetone gradient elution. The method has been applied successfully in the analysis of astaxanthin from X. dendrorhous within less than 20 min, however, the separation of other carotenoids peaks was not satisfactory. Our method described here, in contrast, permits an efficient chromatographic peaks separation of typical carotenoids from X. dendrorhous.

For this purpose, an HPLC method based on a C\(_{30}\) column was developed; it was more effective in separation of structurally related carotenoids than the C\(_{18}\) column. However, several drawbacks occur with this method, such as the high cost of a C\(_{30}\) column and the longer running time. Therefore, this method was usually only used for the study of carotenoids from botanical materials, because the composition of carotenoids in these materials is generally complicated (Su et al., 2002). But most microorganisms produce only a limited number of carotenoids. Our results showed that a C\(_{18}\) column with an optimized mobile phase would enable to achieve enough resolving power in the analysis of microbial samples.

**Table II. Peak identification results.**

<table>
<thead>
<tr>
<th>Peak no.</th>
<th>Name</th>
<th>RRT</th>
<th>(\lambda_{max}) [nm]</th>
<th>(\lambda_{max}) [reported]</th>
<th>APCI-PI (m/z)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>trans-Astaxanthin</td>
<td>1.0</td>
<td>478</td>
<td>478</td>
<td>[M+H]+, 597.3, [M+H−H(_2)O]+,</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>(Weber et al., 2007)</td>
<td>[M+H−2H(_2)O]+</td>
</tr>
<tr>
<td>2</td>
<td>Dehydro-astaxanthin</td>
<td>1.15</td>
<td>469</td>
<td>470</td>
<td>[M+H]+, 595.4, [M+H−H(_2)O]+,</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>(Rao et al., 2005)</td>
<td>[M+H−2H(_2)O]+</td>
</tr>
<tr>
<td>3</td>
<td>DCD</td>
<td>1.39</td>
<td>461, 489, 524</td>
<td>461, 490, 523</td>
<td>[M+H]+, 597.5, [M+H−H(_2)O]+,</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>(An et al., 1999)</td>
<td>[M+H−2H(_2)O]+</td>
</tr>
<tr>
<td>4</td>
<td>cis-Astaxanthin</td>
<td>1.56</td>
<td>367, 469</td>
<td>371, 470 (Yuan and Chen, 1997)</td>
<td>[M+H]+, 597.4, [M+H−H(_2)O]+,</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>[M+H−2H(_2)O]+</td>
</tr>
<tr>
<td>5</td>
<td>Dehydro-DCD</td>
<td>1.68</td>
<td>461, 429</td>
<td>NA</td>
<td>[M+H]+, 595.4, [M+H−H(_2)O]+,</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>[M+H−2H(_2)O]+</td>
</tr>
<tr>
<td>6</td>
<td>Torulene</td>
<td>5.23</td>
<td>464, 483, 515</td>
<td>460, 485, 518</td>
<td>[M+H]+, 534.2, [M+H−H(_2)O]+,</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>(Weber et al., 2007)</td>
<td>[M+H−2H(_2)O]+</td>
</tr>
<tr>
<td>7</td>
<td>(\gamma)-Carotene</td>
<td>5.68</td>
<td>434, 463, 495</td>
<td>430, 460, 489</td>
<td>[M+H]+, 537.4, [M+H−H(_2)O]+,</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>(Weber et al., 2007)</td>
<td>[M+H−2H(_2)O]+</td>
</tr>
<tr>
<td>8</td>
<td>(\beta)-Carotene</td>
<td>6.31</td>
<td>430, 452, 481</td>
<td>425, 452, 478</td>
<td>[M+H]+, 537.4, [M+H−H(_2)O]+,</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>(Weber et al., 2007)</td>
<td>[M+H−2H(_2)O]+</td>
</tr>
</tbody>
</table>

NA, not available in the literature.
There are two carotenoids biosynthetic pathways postulated in *X. dendrorhous*, namely the monocyclic pathway and the dicyclic pathway (An et al., 1999). According to our result, most identified carotenoids were compounds of the monocyclic biosynthetic pathway or their derivatives. It is presumed that the monocyclic biosynthetic pathway may be the dominating biosynthetic pathway in this astaxanthin-overproducing *X. dendrorhous* mutant.

**Acknowledgements**

This research was financially supported by the Hisoar Company and New Century Talents Support Program of the Ministry of Education of China in 2006. We are most grateful to Xiaoman Gu of Analytical and Testing Center, Huazhong University of Science and Technology, Wuhan, China, for HPLC-APCI-MS analyses.