Determination of Hydroxyoctadecadienoic Acids

Nikolay Youhnovski, Daria Schuls, Caroline Schwarz, Gerhard Spiteller, and Klaus Schubert

\* Department of Organic Chemistry 1, Bayreuth University, Universitaetstr. 30, 95447 Bayreuth, Germany. Fax: 0049/921/552671.
\* E-mail: gerhard.spiteller@uni-bayreuth.de

b Institute for Clinical Chemistry and Laboratory Diagnosis of the Friedrich Schiller University Jena, D-07740 Jena, Germany

\* Author for correspondence and reprint requests

Z. Naturforsch. 58c, 268–276 (2003); received October 16/November 14, 2002

Oxidation of low-density lipoproteins (LDL) plays a crucial role in inflammatory diseases and aging. The main oxidation products of LDL are stereoisomeric 9-hydroxy-10,12-octadecadienoic acids (9-HODEs) and 13-hydroxy-9,11-octadecadienoic acids (13-HODEs). Nevertheless the content of HODEs in natural oxidized LDL is low compared to other components, thus determination of HODEs requires a sample enrichment in most cases. Big losses are encountered during the necessary processing due to the instability of HODEs against acidic conditions. Therefore the use of labeled standards is required. Standards with an $^{18}\text{O}$ label in the carboxylic group used previously may partly suffer a loss of the label by exchange with water. In this paper we describe an improved work-up procedure and the preparation of standards labeled with $^{18}\text{O}$ in the hydroxylic group which is not exchangeable.

Key words: Lipid Peroxidation, Hydroxyoctadecadienoic Acids, Inflammatory Diseases

Introduction

Lipid peroxidation (LPO) of polyunsaturated fatty acids (PUFAs) plays a dominant role in the genesis of inflammatory diseases, such as atherosclerosis (Lusis, 2000). Moreover, LPO processes are involved in the deterioration of fat by storage (Frankel 1998), since PUFAs are extremely sensitive to oxidation by air (for a recent review see Spiteller 2002). The most abundant PUFA in tissue and blood of mammals is linoleic acid (Esterbauer et al., 1989) Its peroxidation generates stereoisomeric 9-hydroperoxy-10,12-octadecadienoic acids (9-HPODEs) and 13-hydroperoxy-9,11-octadecadienoic acids (13-HPODEs). These lipiddydroperoxides (LOOHs) are readily reduced in biological surrounding (Wang and Powell, 1991; Wang et al., 1992) to the corresponding alcohols, (LOHs), 9-hydroxy-10,12-octadecadienoic acids (9-HODEs) and 13-hydroxy-9,11-octadecadienoic acids (13-HODEs) as outlined in Scheme 1.

Since HODEs are the most abundant LPO products in blood their determination allows insight into the extend of LPO processes. Nevertheless HODEs are present – compared to other blood and tissue components – even in severe diseased states – only in low amounts. Thus their detection requires the use of sensitive and specific identification methods and also usually the application of enrichment procedures.

Sensitive immunological methods, based on antibodies raised against oxidatively damaged LDL have been found to be not specific enough (Esterbauer et al., 1992). A fast and simple method for determination of all LOHs and LOOHs – in free form or esterified – uses the typical UV absorption of the diene system adjacent to a CH(OH) or a CH(OOH) group at 234 nm. Since compounds with conjugated double bonds are rare in nature (exceptions are trans fatty acids) (Zock and Katan, 1997) quantification of oxidized LDL samples...
is achieved by separation on reversed phase HPLC (Ahotupo and Vasankari, 1999). Although this procedure is fast, it suffers from the missing ability to detect and determine single components. The separation in single components requires a second HPLC step using normal phase columns (Wu et al., 1995).

Alternatively LOHs are determined by gas chromatography combined with mass spectrometry (GC/MS). This method needs sample enrichment. Enrichment is achieved by chromatography on silica gel originally applied to investigate hydroxy derivatives of arachidonic acid by GC/MS (Hamberg and Samuelsson, 1967). This step is combined with sample losses due to adsorption of free HODEs on silica gel. Therefore solid phase extraction on C-18 cartridges is recommended (Powell, 1982).

The isolated fraction of hydroxy acids is then derivatized to transform LOHs to volatile derivatives – necessary for separation by GC: The carboxylic groups are either protected by methylation or transformed to their pentafluorbenzylesters (Strife and Murphy, 1984). Finally the OH groups are subjected to trimethylsilylation (Boeynaems et al., 1980; Aisen et al., 1985; Wang et al., 1992; Lehmann et al., 1992). Thus generated samples are separated by GC and characterized either by registration of electron impact (EI) mass spectra (MS) (Wang and Powell, 1991; Wang et al., 1992) or negative ion (NI) respectively chemical ionization (CI) mass spectra (Strife and Murphy, 1984). Since the EI/MS of thus derivatized saturated hydroxy fatty acids are more specific than those of LOHs usually a hydrogenation step is included before derivatization (Hamberg and Samuelsson, 1967; Lehmann et al., 1992).

In this procedure standards are used for quantification, for instance, saturated or unsaturated hydroxy acids – not occurring in the biological material (Wang et al., 1992; Jira et al., 1996), – but better is the application of hydroxy acids labeled with isotopes. Usually labeled HODEs are prepared by exchange of the carboxylic oxygen by $^{18}$O (Strife and Murphy, 1984). Nevertheless this method was also criticized since $^{18}$O/$^{16}$O exchange occurs in biological media (Gleispach et al., 1985).

Considering these difficulties we exchanged the standards labeled with $^{18}$O in the carboxylic group by HODE standards labeled with $^{18}$O in the hydroxy groups. By checking these standards we recognized severe sample losses during the different steps of sample processing. In this paper we describe the preparation of the new standard and report on a separation procedure which minimizes the losses of HODEs.

**Experimental**

**Materials, chemicals, and reagents**

HCl aqueous solution 32% (puriss p.a.), MeOH, hexane, 2-propanol, THF, CHCl$_3$ (HPLC-grade), ethyl acetate, HOAc, NaOH, KOH and SnCl$_2$ were obtained from Merck (Darmstadt, Germany); PtO$_2$, linoleic acid (99% GC), BHT (= 2,6-bis-tertiary-butyl-4-methyl-phenol), Na$_2$HPO$_4$, NaH$_2$PO$_4$, heparin sodium salt (porcine-intestinal mucosa), and lipoxidase = lipoxygenase (soybean, 12.4 U/mg) were bought from Fluka (Buchs, Switzerland). 5% rhodium alumina powder was obtained from Aldrich (sample number: 21, 285–7). The $^{18}$O$_2$ (99% $^{18}$O isotope) was purchased from Linde AG (Unterschleissheim, Germany). Nitrogen, argon, and oxygen were from Riessner Gase (Lichtenfels, Germany). The H$_2$O was purified with an MILLI-RO apparatus (Millipore, Milford, MA, USA).

**Equipment**


GC: HRGC Model 416 (Carlo Erba Strumentazione, Milano, Italy), carrier gas: hydrogen (pressure 0.5 bar); capillary fused silica DB-5 column J&C Scientific (Mainz-Kastel, Germany) length 30 m; splitting: 1:30; injection temperature: 270 °C. FID detector temperature: 290 °C.

GC-MS: Hewlett-Packard 5890 Series II; carrier gas: hydrogen (pressure 0.5–0.8 bar); no splitting; capillary column: fused silica DB-5 (J&C Scientific), length: 30 m, i.d. 0.25 mm; film thickness: 0.1–0.4 µm; injection temperature 270 °C. Finnigan MAT 95 mass spectrometer with inverse Nier-Johnson geometry and EI-CI ion source. ESI-MS: Finnigan TSQ 700 mass spectrometer, equipped with a Finnigan electrospray ionization (ESI) ion source.
Chromatographic conditions

Reversed phase preparative HPLC:

LiChrosorb RP-18 (250 × 25 mm, 7 µm, Merck) C18 column; mobile phase: isocratic: H2O-MeOH (1:9) and 0.2% HOAc; flow 20 ml/min; detection at 234 nm.

Reversed phase analytical HPLC:

The attempts to separate 9- and 13-HODE on different endcapped and not endcapped C18, C8 and C2 columns with different polar mobile phases were not successful, but separation was achieved using a ET 250/4 Nucleosil 100-5 C4 column (Macherey-Nagel) and as gradient mobile phase: H2O-MeOH 0.2% aqueous solution of HOAc, gradient: 0 min – (30:60:10, v/v) → 50 min – (5:85:10); flow rate 1 ml/min, injection volume 10 µl, detection at 234 nm. Validation: the calibration curves show linearity in the interval 30 ng – 3 µg/l. The differences for back calculated concentrations did not exceed 5% on average. The detection limit was 5 ng at a signal to noise ratio 3:1.

Normal phase preparative HPLC:

(Wu and Robinson, 1995)

Ultrasep FS 100 (250 × 25 mm, 6 µm) column Bischoff (Leonberg, Germany); mobile phase: isocratic: hexane-2-propanol-HOAc (986:12:2, v/v); flow: 20 ml/min; detection: 234 nm.

GC-MS:

Hewlett-Packard 5890 Series II; carrier gas: hydrogen (pressure 0.5–0.8 bar); no splitting; capillary column fused silica DB-5 (J&C Scientific), length: 30 m, i.d. 0.25 mm; film thickness: 0.1–0.4 µm; injection temperature 270 ºC. Finnigan MAT 95 mass spectrometer with inverse Nier-Johnson geometry and EI-CI ion source. Temperature gradient: 120 ºC → 300 ºC with 3 ºC/min.

Mass spectrometry

GC-MS:

Carrier gas: hydrogen (pressure 0.5–0.8 bar); no splitting; capillary column fused silica DB-5 (J&C Scientific), length: 30 m, i.d. 0.25 mm; film thickness: 0.1–0.4 µm; injector temperature 270 ºC.

Standard preparation

18O (9S, 10E, 12Z)-9-hydroxy-10,12-octadecadienoic acid and 18O (13S, 9Z, 11E)-9-hydroxy-9,11-octadecadienoic acid (analogous to enzymatic preparation of HP16ODEs, (Gardner, 1989):

A solution of 0.3 g linoleic acid in 90 ml phosphate buffer (0.1 m, pH 8) was filled in a three-neck flask, the air was removed by evacuation. The flask was ventilated three times with Ar. 40 mg soybean lipoxidase = lipoxygenase (12.4 U/mg) were added under a constant Ar flow. After additional evacuation and Ar ventilation the reaction system was filled with 18O2. The solution was stirred for 1 h at 25 ºC. An additional portion of 18O2 was added and the reaction mixture was stirred for another 1 h. The closed system was opened and 1.5 g NaBH4 were carefully added (foam) under Ar ventilation. After 2 h the reaction mixture was acidified with 1 m aqueous solution of HCl (pH 3–3.5), saturated with NaCl, extracted three times with 100 ml CHCl3-MeOH (3:1) and finally two times with 75 ml CHCl3. In order to avoid losses of the product during the solvent evaporation 10 ml H2O were added (see discussion). The solvents were removed in vacuum (temperature below 30 ºC). The obtained colorless oil was separated by preparative normal phase HPLC: Ultrasep FS 100 (250 × 25 mm, 6 µm) column (Bischoff, Leonberg, Germany); mobile phase: isocratic hexane-2-propanol-HOAc (986:12:2, v/v); flow: 20 ml/min; detection: 234 nm. The isotopic purity of the obtained 9-HP18ODE and 13-HP18ODE was determined by ESI-MS to be 95%. 9-H18ODE and 13-H18ODE were used as internal standards for quantification. Yield: 40 and 35 mg respectively.

LDL sample preparation and work up procedure

Fresh blood samples (10 ml) from patients were transferred in ice to the laboratory. After coagulation (ca 30 min, 4 ºC) the sample was centrifuged for 10 min at 9300 × g/min. The LDL-fraction was isolated according to the method of Leiß (Leiß et al., 1979): briefly the precipitating reagent 1 – in an amount of 10% v/v in respect to the serum volume – was added to the separated blood serum (30–40% v/v of the blood sample). The mixed sample was kept for 1 h in the dark (ice bath) and centrifuged (30 min, 4 ºC, 20,900 × g). The pellet
(VLDL) was separated, then the supernatant was mixed with the precipitating reagent 2, cooled for 1 h in the dark (ice bath) and centrifuged (30 min, 4 °C, 20,900 × g). The pellet (LDL) was stored at −20 °C under Ar.

The LDL pellet was dissolved in a mixture of 20 ml phosphate buffer (0.1 m, pH 7.4) and 100 µl BHT solution. Then 10 µl of the standard solution, containing an exactly weighted amount of 10 µg of 9-HODE and 13-HODE labeled with 18O in the hydroxyl group was added. The pellet was dissolved by stirring applying ultrasound. The solution was transferred to a funnel, 75 ml of a solution consisting of CHCl₃-MeOH (1:2) were added. The mixture was shaken. Then 5 ml CHCl₃ were added, the solution was shaken again. 20 ml of a 0.5 m NaH₂PO₄ solution were added and shaken again. The solution separated into two layers. The water layer was extracted three times with 50 ml CHCl₃. The collected organic extracts were evaporated in vacuum (room temperature). The residue was weighted (crude LDL, 5–30 mg).

Lipid hydrolysis: the obtained crude LDL was dissolved in 6 ml 1 m KOH/MeOH-H₂O (9:1) solution and stirred under Ar for 1.5 h at 50 °C (Ulberth and Kamptner, 1992). The cooled solvent was neutralized with 24.5 ml 1 m aqueous solution of mono-sodium dihydrogen phosphate (pH 5.8–6). 64 ml CHCl₃ and 58 ml MeOH were added to the neutralized solution. The mixture was shaken. Layer separation was achieved by addition of 32 ml H₂O. The H₂O phase was extracted additionally three times with 50 ml CHCl₃. ~ 10% H₂O v/v was added to the collected extracts and the mixture was evaporated in vacuum keeping the temperature below 30 °C. The residue was dissolved in 3 ml CHCl₃ and loaded into a homemade appropriately equilibrated SPE cartridge. The cartridge was washed with 4 ml CHCl₃. The acidic fraction was eluted with 4 ml 2% HOAc in CHCl₃ and flushed with nitrogen. Solvents were removed from the eluate by continuous nitrogen flow without heating. The samples were then dissolved in 0.5 ml CHCl₃ and derivatized by addition of 1 ml 4% solution of diazomethane in Et₂O, containing 5% of methanol. Then the samples were dissolved in AcOEt, about 10 mg of a 5% Rh catalyst on alumina powder were added and the samples were hydrogenated at 1 bar H₂ pressure for 10 min (Wang et al., 1992). After removal of the catalyst and the solvent unsubstituted methylesters were separated from hydroxylated ones by silicagel flash chromatography. The residue obtained after hydrogenation was weighted again in order to determine the amount of HODE in LDL. Then the sample was dissolved in 0.5 ml CHCl₃ and treated with MSTFA (100 µl) for 1 h at 40 °C. The solvents were removed by continuous nitrogen flow. The dried samples were dissolved in a definite volume CHCl₃ (concentration 10 mg/ml). An equivalent of the solution was injected into the GC-MS for quantification.

Preparation of precipitation reagent 1:

5 ml solution of heparin sodium salt (2500 IU/ml) are mixed with 5 ml 2 m aqueous solution of MgCl₂ (Leiß et al., 1979).

Preparation of precipitation reagent 2:

5 ml 2m aqueous solution of MgCl₂ are mixed with 5 ml aqueous solution of dextrane sulfate sodium salt 20 g/l (Leiß et al., 1979).

Preparation of the home-made SPE cartridges:

A 2 ml glass syringe was filled with 0.4 g amino-propyl-modified silica gel CROMABOND LV-NH₂ and filter elements from filter paper 595 Schleicher & Schüll GmbH (Dassel, Germany). The packed cartridges were equilibrated with 4 ml hexane and 1 ml CHCl₃.

Esterification with diazomethane (esterification of -COOH groups) (Hamberg and Fahlstadius, 1990).

The samples are dissolved in 0.5 ml CHCl₃ and 1 drop of methanol (addition of methanol traces accelerate the reaction, private communication by Dr. Gardner), then 1 ml 4% solution of diazomethane in Et₂O is added. After 3 min the solvents are removed by a continuous nitrogen flow.

Catalytic hydrogenation of the double bonds analogous to Wang (Wang et al., 1992).

The samples are dissolved in 6 ml methanol. After addition of 5 mg 5% rhodium alumina powder the suspension is shaken under hydrogen (pressure 1 bar) for 10 min. The catalyst is filtered off and the solvent is evaporated (continuous nitrogen flow).
Separation of hydroxy acids from unsubstituted acids:

The residue obtained after hydrogenation is dissolved in 2 ml cyclohexane-EtOAc (98:2). This solution is brought onto a silicagel column (5 g) equilibrated with cyclohexane-EtOAc (98:2). The column is washed with 80 ml of the same solvent mixture. Hydroxylated methylates are eluted by rinsing the column with 80 ml EtOAc.

Trimethylsilylation with N-methyl-N-trimethylsilyl-trifluoroacetamide (MSTFA, protection of hydroxy groups)

The samples obtained after removal of the solvent are dissolved in 0.5 ml CHCl₃ and heated with MSTFA (100 µl) for 1 h (40 °C). The solvents are removed by a continuous nitrogen flow.

Blank recovery experiments
Blank experiment 1:

In order to check the loss of HODEs (by application of previous separation procedures) 20 µg 9- and 13-HODE are dissolved in a mixture of 20 ml phosphate buffer (0.1 m, pH 7.4) and 200 µl BHT solution (2% in ethanol). 100 ml CHCl₃-MeOH (v/v) are added and the mixture is shaken. Separation in two layers is achieved by addition of 23 ml H₂O and 2 ml 1 m aqueous solution of HCl. The organic layer is separated and the water layer is extracted three times with 50 ml CHCl₃. The collected extracts are evaporated in vacuo. The residue is dissolved in a mixture of 1 ml THF and 2 ml 1 m NaOMe methanolic solution and stirred under Ar 1 h at room temperature. The solvent is acidified with 1 m aqueous solution of HCl (pH 2). The THF is evaporated and the residue is dissolved in 50 ml water and extracted three times with 50 ml CHCl₃. The collected extracts are dried with Na₂SO₄ and then subjected to evaporation in vacuum. Then the residue is taken up in CH₃OH, hydrogenated (as described above but using PtO₂ as catalyst) and derivatized with diazomethane and MSTFA. The dried samples are dissolved in a definite volume CHCl₃ and are analyzed by GC-MS. 97% ± 2 (average value from 10 samples) of HODEs were transformed to stearic acid.

Blank experiment 2:

The same HODE samples are processed as described above but the collected CHCl₃ extracts are not dried. The HODE losses are found to amount to 90–95% (average of ten samples).

Blank experiment 3:

is carried out like blank experiment 2, but instead 1 N HCl, a 1 m aqueous solution of NaH₂PO₄ is used for neutralization and the catalyst is substituted by rhodium on alumina. The losses were found to be less than 10% (average of ten samples).

Hydrogenation experiments

Hydrogenation experiment 1:

0.8 mg PtO₂ catalyst are added to 48 µg of 13-HODE dissolved in 4 ml of methanol. 5 ml of CHCl₃ are added. The solution is shaken for 30 min in a H₂ atmosphere at a pressure of 1 bar. The products are investigated after trimethylsilylation by GC-MS. Two peaks of equal height are recognized. The first corresponds to the TMS derivative of 13-hydroxystearic acid, the second to stearic acid.

Hydrogenation experiment 2:

Repeating of the experiment by hydrogenation in 10 ml AcOEt instead of CH₃OH-CHCl₃ results in an improvement of the yield (ratio of 13-hydroxystearic acid to stearic acid 4:1).

Hydrogenation experiment 3:

In a next experiment hydrogenation in CH₃OH-CHCl₃ is carried out with the methylate of 13-HODE. The yield improves considerably compared to hydrogenation of the free acid (ratio 13-hydroxystearic acid methylate to stearic acid methylate 3:1). 13-Methoxystearic acid methylate is recognized as byproduct.

Hydrogenation experiment 4:

Hydrogenation of the methylate of 13-HODE by exchange of PtO₂ versus 5% rhodium alumina powder as catalyst in AcOEt yields only traces of stearic acid methylate, but results also in partial transformation of the hydroxylic group to the oxo
group generating 13-oxostearic acid methylate as byproduct (ratio 13-oxostearic acid methylate: trimethylsilylated 13-hydroxystearic acid methylate 1:10).

**Results and Discussion**

**Establishment of the extraction method**

Already nearly four decades ago it has been recognized that homogenation of mammalian (Wills, 1966) or plant (Galliard, 1970) tissue in aqueous solutions – even at 0 °C (Galliard, 1970) – is inevitably connected with massive LPO processes. Thus large amounts of HODEs are generated artificially and consequently determination of LPO products after homogenation of tissue in aqueous solvents does not reflect the original extend of LPO processes in physiological and diseased states. Nevertheless often homogenation is still carried out in buffer solutions (Sohal et al., 1994; Wu et al., 1995). This artificial generation of LPO products can be prevented substantially when enzymes are destroyed before homogenation by boiling of the sample or by paralyzing their action using for extraction an organic solvent. In spite of these precautions a moderate generation of LPO products can not be excluded completely by processing of tissue samples due to their high water content combined with the immediately activation of enzymes by cell injury which might allow to induce LPO for a short time before the enzymes are inactivated. More reliable results are obtained if tissue samples are subjected to deep freezing (Lehmann et al., 1992). Nevertheless already the removal of organs might be connected with partial injury and thus induction of LPO processes. The danger to initiate LPO processes is much lower when biological liquids, e.g. blood or liquor, are analyzed because mechanical destruction is avoided.

As mentioned above, unambiguous identification of HODEs requires a GC/MS analysis after appropriate enrichment and derivatization (Aisen et al., 1985; Kühn et al., 1989; Wang and Powell, 1991; Lehmann et al., 1992; Jira et al., 1996). When we checked the validity of the previously widely applied methods by use of 18O labelled standards (see below) we observed great sample losses. Therefore we investigated each step of the sample processing by GC/MS analysis. Thus we recognized that predominantly traces of acid are responsible for loss of samples:

LOHs are secondary allyl alcohols, their sensitivity to acids is well known. Therefore LOH esters are saponified usually in alkaline media. The separation of the acids from the neutral compounds requires acidification. In previous investigations samples have been acidified to a pH 2 (Wu et al., 1995; Jira et al., 1996) or even to pH 3.1–3.2 either by addition of HCl (Gleispach et al., 1985) or an ammonium formate buffer (Lehmann et al., 1992). We observed extreme sample losses by acidifying the organic extract with HCl to pH 3, especially when the extracts were dried before solvent removal: HCl and other acids are transferred partly into the organic layer. If the organic extract is dried acid is accumulated in the residue. The acid catalyzes elimination of water, transferring HODEs to a mixture of trienoic acids which are transformed to steric acid by catalytic hydrogenation. The latter is easily detectable after derivatization by GC/MS. The yield of HODEs is improved if the organic solvents are not dried because then acids are removed by azeotropic distillation when the organic solvent is distilled off. Therefore addition of 10–15% (v/v) of water to the organic solvents before distillation prevents at least partly the decomposition of LOHs.

The observation that weakly acidified solutions induce water elimination of HODEs is also important for their separation on normal phase HPLC columns (necessary for instance for standard preparation) where separations require addition of acetic acid (various amounts of acetic acid between 6% (Gleispach et al., 1985) and 0.001% (Clare et al., 1991) have been recommended). The lower the amount of acetic acid added the lower are the losses.

Nevertheless accumulation of acetic acid by removal of solvent can not be avoided completely. Therefore we neutralized the basic solutions obtained after saponification with a 1 M NaH₂PO₄ solution only to pH 5.8. This improved the yield of HODEs dramatically.

The fraction of acids is then separated from unpolar products by SPE (Solid Phase Extraction) on a weekly basic ion exchange column (amino-propyl-modified silica gel). Since elution of acids requires addition of 2% of acetic acid to the chloroform solution, evaporation of the solvent must
be carried out carefully on a water bath not exceeding a temperature of 30 °C and after addition of approximately 10% of water (necessary for azeotropic removal of acid).

The tendency for water elimination in further processing is reduced if the double bonds are hydrogenated. Moreover derivatives of saturated hydroxy acids show very characteristic electron impact mass spectrometric fragmentation patterns (Lehmann et al., 1992; Wu et al., 1995).

The hydrogenation experiments revealed that hydrogenation of free HODEs by use of PtO2 as catalyst and methanol as solvent generated large amounts of stearic acid, the sample losses were reduced when the free acids were converted to their methylates and even more by exchange of the solvent to ethylacetate. Best results were obtained by exchange of the PtO2 catalyst by a 5% rhodium catalyst on alumina powder as recommended by Wang (Wang et al., 1992) and reducing the time of hydrogenation. Nevertheless we recognized also in this case a side reaction, the generation of saturated keto-acids.

The exchange of the OH group versus hydrogen observed by catalytic hydrogenation is thus apparently caused by a slightly acidic solvent and prolonged exposure to too large amounts of catalyst in respect to the sample amounts.

Polar groups cause partial sample decomposition on GC columns, moreover hydroxy-compounds are partly adsorbed. Therefore the free hydroxygroups must be converted to their trimethylsilylethers by treatment with N-methyl-N-trimethylsilyl-trifluoroacetamide (MSTFA) or another silylating reagent before GC/MS investigation of the samples is started.

Although samples are ready for GC/MS analysis after derivatization it might be sometimes useful to remove the main amount of less polar saturated fatty acid methylates by flash chromatography on silicagel. Since free acids are adsorbed on silicagel columns to a much higher extend than methylates also this step requires preparation of esters.

### Choice of the internal standards

The inevitable losses in the multistep procedure of sample preparation for quantification of HODEs require the application of internal standards. When saturated hydroxyacids of unusual chain length are used for this purpose (Wang and Powell, 1991; Jira et al., 1996) losses occurring due to water elimination of HODEs and during hydrogenation escape partly detection. Better internal standards for quantification of HODEs are isotopically labeled analogues of HODEs. In previous investigations hydroxy acids derived by LPO processes were labeled at the carboxyl group with 18O (Strife and Murphy, 1984), but exact quantification might be aggravated due to partial exchange of the 18O to 16O in biological media (Gleispach et al., 1985). In contrast exchange of the 18O labeled oxygen atom in the hydroxy group of secondary alcohols versus 16O is impossible under normal circumstances. Therefore we synthesized H18ODEs in analogous manner as described for H16ODEs (Gardner, 1989) by reacting linoleic acid with soybean lipoxygenase in an 18O2 atmosphere. The ob-

<table>
<thead>
<tr>
<th>9.16O-HODE</th>
<th>9.18HODE</th>
<th>m/z 229/231</th>
<th>Number of experiments</th>
<th>Experimental m/z 229/231 average peaks ratio</th>
<th>Standard deviation</th>
<th>Error</th>
<th>Lower limit</th>
<th>Upper limit</th>
<th>% RSD</th>
<th>Accuracy</th>
</tr>
</thead>
<tbody>
<tr>
<td>9.16O-HODE as an internal standard</td>
<td>theoretical</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
tained $[^{18}\text{O}]-\text{HODEs}$ were determined to have an isotope purity greater than 95% (by ESI/MS). HODEs proved to be stable if stored without solvent in an argon atmosphere.

**Determination of LDL**

An aliquot of LDL samples processed as described in the experimental section was dissolved in MSTFA and then subjected to separation by GC/MS.

GC peaks corresponding to derivatized 9- and 13-hydroxystearic acid and their $^{18}\text{O}$ labeled isomers were recognized by their typical EI/MS spectra. Quantification was achieved by measuring the ion currents of the $\alpha$-fragments as described (Wang and Powell, 1992; Lehmann et al., 1995) by registering the ion currents of the $\alpha$-fragments found at $m/z$ 173 and $m/z$ 315 in the not labeled and at $m/z$ 175 resp. 317 in the labeled 13-HODE spectra (Fig. 1). Likewise the fragments of 9-HODE were registered at $m/z$ 229 and $m/z$ 259, resp. at $m/z$ 231 and $m/z$ 261. The ion currents of these fragments can be measured with great accuracy (see Table I).

**Conclusions**

The reported improved processing for HODE quantification minimizes the sample losses during HODE extraction from biological samples caused by presence of strong acids. SPE is a quick and easy method for preliminary separation of the acidic sample fraction from the matrix and allows to increase the sensitivity. Quantification was achieved with $^{18}\text{O}$ isotopically labeled internal standards, containing the label in the hydroxylic group. Application of the method will be published elsewhere.

![Fig. 1. Standard curves for (A) 9-HODE and (B) 13-HODE. Isotopically labeled 9-\(^{18}\text{O}\)HODE and 13-\(^{18}\text{O}\)HODE were used as internal standards (10 µg). 9-HODE was monitored by measuring the ion intensities at $m/z$ 229 (A) and $m/z$ 259 (data not shown), 13-HODE was monitored by measuring the ion intensities at $m/z$ 173 (B) and $m/z$ 315 (data not shown). The internal standard: 9-\(^{18}\text{O}\)HODE was monitored by measuring the ion intensities at $m/z$ 231 and $m/z$ 261 and 13-\(^{18}\text{O}\)HODE was monitored by measuring the ion intensities at $m/z$ 175 and $m/z$ 317. Ion current peaks at the appropriate GC retention times were integrated. The ratios of the peak integrals reflecting the target analyte divided by that of the internal standard were plotted as a function of the amounts of (A) 9-HODE and (B) 13-HODE.](image)

**Acknowledgement**

We thank Deutsche Forschungsgemeinschaft (DFG), Fonds der Chemischen Industrie and Fischer Stiftung for financial support. We are obliged to Kerstin Hannemann for the careful investigation of the blank hydrogenation experiments and to Mr. M. Glaessner for mass spectrometric measurements.


Spiteller G. (2002), Do changes in the cell membrane structure induce the generation of lipid peroxygenation products which serve as first signalling molecules in cell to cell communication? Prostaglandins, Leukotrienes & Essential Fatty Acids 67, 151–162.


