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# Isolation and Identification of Lathycarpin, a New Pterocarpan Phytoalexin from Lathyrus sativus

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After treatment with the fungus *Helminthosporium car-bonum* or aqueous copper sulphate, excised *Lathyrus sati-vus* leaflets produce pisatin and a second isoflavonoid phytoalexin (lathycarpin) identified as (+)-6aR; 11aR-2,3-di-methoxy-6a-hydroxy-8,9-methylenedioxypterocarpan.

# Introduction

Earlier studies [1, 2] have demonstrated that a number of isoflavonoid (pterocarpan) phytoalexins [3] accumulate in excised, fungus-inoculated leaflets of species belonging to the genus *Lathyrus* (Leguminosae-Papilionoideae; tribe Vicieae). Apart from pisatin (3-methoxy-6a-hydroxy-8,9-methylenedioxypterocarpan, 1), the most commonly encountered *Lathyrus* phytoalexin, small quantities of three other fungitoxic pterocarpans (medicarpin, maackiain and variabilin) are variously produced by several *Lathyrus* spp. [2]. In addition, nissolin and its 9-O-methyl ether (methylnissolin) are formed, to-

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gether with medicarpin, by fungus-treated phyllodes of L. nissolia although as yet these two unusual 3,9,10-trisubstituted pterocarpans have not been discovered elsewhere in the genus [4]. Pisatin and the non-isoflavonoid chromones, lathodoratin methyl-lathodoratin, occur as phytoalexins in tissues of L. odoratus (sweet pea) [5], whilst some evidence has been obtained to suggest that the Indian pulse L. sativus (grass pea; chickling pea) produces 1, and a new pterocarpan (designated LS-2) of undetermined structure [1]. We have now re-examined the phytoalexin response of L. sativus and can confirm the ability of detached leaflets to accumulate 1 following treatment with a spore suspension of Helminthosporium carbonum [1, 2]. This fungus also stimulates the formation of a second antifungal isoflavonoid, probably identical with LS-2, which we propose to name lathycarpin. The characterisation of lathycarpin (+)-6aR; 11aR-2,3-dimethoxy-6a-hydroxy-8,9methylenedioxypterocarpan (2) is described in this report.

# **Results and Discussion**

Pisatin and lathycarpin were initially isolated from H. carbonum-inoculated L. sativus leaflets (but not from those treated with de-ionised H<sub>2</sub>O) using the drop-diffusate technique outlined in the Experimental section. This procedure was very tedious, however, and during later stages of the project was replaced by a modified diffusion method (see Experimental) in which leaflets were floated for 10-12 days on aqueous CuSO<sub>4</sub>. After this period, the CuSO<sub>4</sub> solution was shaken with EtOAc to remove all isoflavonoid compounds, subsequent Si gel TLC purification of the organic phase yielding both 1 and 2 in milligram quantities. Pisatin was identified by UV, MS and Si gel TLC comparison with authentic material obtained from Pisum sativum [6]. Other compounds currently recognised as Lathyrus phyto alexins (see Introduction) were not produced by L. sativus.

The UV (EtOH) spectrum of chromatographically pure lathycarpin ( $\lambda$  max: 212, 235 sh, 303 nm) closely resembled that of 2,3-dimethoxy-8,9-methylenedioxypterocarpan ( $\lambda$  max: 212, 235 sh, 304 nm), and could be distinguished from those of spectroscopically similar 3,4-dioxygenated pterocarpans (e.g. 4-methoxymaackiain) by the absence of two



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slight shoulders between 270 and 290 nm [7]. Aqueous NaOH had no effect on the neutral UV spectrum of lathycarpin, whereas addition of conc. HCl resulted in rapid (1 min) dehydration to afford the corresponding pterocarpene ( $\lambda$  max: 353 and 368 nm). This acid-mediated spectroscopic change is characteristic of pterocarpans (e.g. pisatin and variabilin) containing a labile C-6a OH group [7, 8]. Further evidence for tertiary hydroxylation of 2 (High Resolution MS, M<sup>+</sup> 344.0892;  $C_{18}H_{16}O_{7}$ ; 55%) was provided by the mass spectrum which exhibited a major fragment at m/z 326.0803 (M<sup>+</sup>-H<sub>2</sub>O; 100%) corresponding to  $C_{18}H_{14}O_6$  (cf. 1, M+ 314 (28%), m/z296 (M<sup>+</sup>-H<sub>2</sub>O; 100%)). Other prominent ions in the MS of 2 appeared at m/z 345 (M<sup>+</sup> + 1; 11%), 329  $(M^+-CH_3; 5\%), 327 (22\%), 325 (31\%), 316 (16\%),$ 311 (16%), 301 (6%), 285 (13%), 283 (7%), 199 (9%), 163 (26%), 151 (9%), 149 (14%) and 133 (10%). On TLC plates sprayed with chromotropic acid reagent, lathycarpin gave a purple-pink colour indicative of methylenedioxy substitution [9, 10].

In view of its co-occurrence with 1, the above data suggested that lathycarpin was a methoxy-pisatin, and this was confirmed by <sup>1</sup>H NMR spectroscopy (Table I) which revealed that pisatin and lathycarpin differed only in that the latter compound possessed an extra methoxyl substituent, and lacked a C-2 proton. The new phytoalexin is thus 2-methoxypisa-

Table I. <sup>1</sup>H NMR data for pisatin and lathycarpin <sup>a</sup>.

Proton	Pisatin (1)	Lathycarpin (2)
H-1	7.36 d, 1H ( <i>J</i> = 8.5 Hz)	6.96 s, 1H
H-2	$6.63 \mathrm{q}, 1 \mathrm{H}$ ( $J = 8.5, 2.5 \mathrm{Hz}$ )	-
H-4	$6.39 \mathrm{d}, 1 \mathrm{H}$ ( $J = 2.5 \mathrm{Hz}$ )	6.43 s, 1H
H-6ax/eq	4.11 s, 2H	$4.06 \mathrm{d},  2\mathrm{H}$ ( $J = 1.5 \mathrm{Hz}$ )
H-7	6.89 s, 1H	6.89 s, 1H
H-10	6.35 s, 1H	6.36 s, 1H
H-11a	5.29 s, 1H	5.26 s, 1H
$O-CH_2-O$	$5.93 \mathrm{q}, 2 \mathrm{H}$ ( $J = 2.9, 0.9 \mathrm{Hz}$ )	$5.92 \mathrm{q},  2\mathrm{H}$ ( $J = 2.9,  0.9 \mathrm{Hz}$ )
OCH3	3.75 s, 3H	{ 3.76 s, 3H 3.7 8s, 3H

<sup>&</sup>lt;sup>a</sup> Solvent,  $(CD_3)_2CO$ ; chemical shifts are given as  $\delta$  values (TMS reference); figures in parentheses refer to coupling constants.

tin (2,3-dimethoxy-6a-hydroxy-8,9-methylenedioxypterocarpan, 2), a structure supported by the loss, relative to pisatin, of significant coupling in the H-1 and H-4 signals and by the upfield shift of H-1 consequent upon *ortho*-oxygenation [11].

Lathycarpin has  $[\alpha]_{589 \text{ nm}}^{21} + 232^{\circ}$  (1.2 mg in 1 ml MeOH) and can therefore be assigned the 6aR; 11aR absolute configuration depicted in **2** [12]. Pisatin from *L. sativus* is also dextrorotatory,  $[\alpha]_{589 \text{ nm}}^{21} + 292^{\circ}$  (1.05 mg in 1 ml MeOH).

In TLC plate bioassays [13, 14], lathycarpin (20 µg) was clearly inhibitory to the spore germination/germ tube growth of *Cladosporium herbarum*. More precise measurements against radial mycelial growth of H. carbonum [14] gave an ED<sub>50</sub> value of about 45 µg/ml, comparable with that reported earlier for pisatin [15]. On average, diffusates from excised L. sativus leaflets treated with a spore suspension of H. carbonum contained pterocarpans 1 and 2 at concentrations of 18 and approx. 30 μg/ml (based on  $\varepsilon = 7244$  at 309 nm for 1 [8]) respectively. Abiotic induction using droplets of aqueous CuSO<sub>4</sub> was less efficient yielding (over four experiments) only  $5-14 \mu g/ml$  of 1, and  $8-20 \mu g/ml$  of 2. Leaf diffusates invariably contained more lathycarpin than pisatin.

#### **Experimental**

Plant material. Seeds of an unnamed variety of Lathyrus sativus L. (supplied by Dr. L. J. G. Van der Maesen, I.C.R.I.S.A.T., Hyderabad, India) were sown in John Innes No. 1 compost, and the resulting plants grown (20 – 24 °C) for about 6 weeks before individual leaflets were removed for treatment with either H. carbonum or aqueous CuSO<sub>4</sub>. At later stages of growth, the plants were routinely deflowered to encourage leaf production.

Isolation and purification of pisatin (1) and lathy-carpin (2). a) Standard drop-diffusate technique. 2-5 droplets of aqueous CuSO<sub>4</sub> (0.25 g/100 ml de-ionised H<sub>2</sub>O, plus 0.5 ml Tween 20 as a wetting agent) or H. carbonum spore suspensions (also containing Tween 20) [16, 17] were placed along the upper surface of excised L. sativus leaflets floating on tap H<sub>2</sub>O. After incubation (20 °C) for 72 h, the droplets (diffusate) were collected, extracted (× 3) with EtOAc, and the pooled organic fractions reduced to dryness (in vacuo, 40 °C). Si gel TLC (Merck, F-254,

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layer thickness 0.25 mm) of the extracts (CHCl<sub>3</sub> -MeOH, 50:1, 12 h equilibration at 20 °C) gave pisatin + lathycarpin as a broad, fluorescence-quenching band  $(R_F 0.67)$ . This band was removed and the pterocarpans eluted (EtOH) prior to separation by Si gel TLC in n-hexane-acetone, 2:1, 1 h equilibration [1] (1,  $R_F$  0.37; 2,  $R_F$  0.28). Final TLC purification of pisatin and lathycarpin was undertaken using n-pentane-Et<sub>2</sub>O-glacial HOAc, 75:25:3, 1 h equilibration  $(1, R_F 0.31; 2, R_F 0.10)$ . b) Modified diffusion technique. In a typical experiment about 600 L. sativus leaflets were cut into short (1-3 cm) sections and floated (10-12 days) on aqueous CuSO<sub>4</sub> (about 21; see above for composition) in transparent-plastic sandwich boxes covered with clear, cling-film food wrap. The leaf material was ultimately discarded, and the CuSO<sub>4</sub> solution shaken (×2) with equal volumes of EtOAc. Si gel TLC of the organic phase was undertaken as described above. Pterocarpan yields ranged from 0.5 – 1.1 mg for pisatin, and from approx. 1.2-1.9 mg for lathycarpin.

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